

An Impact of Culture Conditions on the Expression of KPNAs Proteins in Cancer Cell Lines

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Summary

This report explores the critical but underexplored connection between two-dimensional (2D) cell culture conditions and the dysregulation of importin alpha, a key nuclear transport protein implicated in cancer. While 2D cultures are widely used, their inherent artificiality particularly the unnatural stiffness of plastic substrates and the lack of a three-dimensional (3D) microenvironment fundamentally alters cellular behavior and gene expression. Importin alpha is essential for shuttling proteins, including oncogenes and tumor suppressors, into the nucleus. Its aberrant expression is a known driver of tumor progression. The report argues that the non-physiological mechanical and architectural cues of 2D culture, via mechanotransduction pathways, likely contribute to the dysregulation of importin alpha. Despite a lack of direct, explicit studies on this specific link, the body of evidence on the broader effects of 2D culture and the known role of mechanobiology in nuclear function strongly suggests a significant knowledge gap. Addressing this gap by conducting direct comparative research between 2D and 3D models is crucial to improve the physiological relevance and translational success of in vitro cancer research.

Part 1. Introduction: The Critical Role of Nuclear Transport in Cancer and the Challenge of In Vitro Models

Nuclear-cytoplasmic transport represents a fundamental cellular process, meticulously orchestrating the selective movement of macromolecules, including proteins and RNA, between the nucleus and cytoplasm. This intricate regulation is indispensable for maintaining cellular homeostasis, precisely controlling gene expression, and coordinating diverse cellular responses to various internal and external stimuli.^{www} In the pathological context of cancer, dysregulation of these nuclear transport

pathways, particularly those mediated by importins, is increasingly recognized as a significant contributor to oncogenesis, tumor progression, and the development of therapeutic resistance.^{www} Importin alpha proteins, serving as key adaptor molecules in the classical nuclear import pathway, play a pivotal role in shuttling a vast array of cargo proteins, including both tumor suppressors and oncogenes, into the nucleus where they exert their functions.^{www} Consequently, a comprehensive understanding of how their expression and functional activity are modulated in different experimental settings is paramount for advancing accurate cancer research and developing effective therapeutic strategies.

For decades, traditional two-dimensional (2D) cell culture models have served as the cornerstone of in vitro cancer research due to their inherent simplicity, cost-effectiveness, and ease of manipulation.^{www} However, the inherent artificiality of these flat, rigid environments, which profoundly differ from the complex three-dimensional (3D) in vivo tumor microenvironment (TME), raises critical questions about the physiological relevance and translational applicability of findings derived from them.^{www} These disparities are not merely superficial; they can lead to significant alterations in fundamental cellular characteristics such as morphology, polarity, cell-cell and cell-extracellular matrix (ECM) interactions, and the availability of nutrients and oxygen.^{www} All these factors are known to profoundly influence global gene expression, protein activity, and overall cellular behavior.^{www}

The profound simplification inherent in 2D culture models, rather than merely failing to replicate in vivo conditions, actively induces aberrant cellular behaviors and gene expression patterns. This suggests that 2D conditions are not inert platforms but rather active contributors to a non-physiological cellular state, which can manifest as dysregulation of critical cellular processes, including those involving importin alpha. This fundamental disconnect between 2D in vitro models and the in vivo reality has significant implications, particularly for drug discovery. If 2D culture conditions lead to a non-physiological expression or function of importin

alpha, or other critical cellular processes, then therapeutic strategies developed and screened in these simplified models might not accurately predict efficacy or toxicity in patients. For example, 3D cultures have demonstrated higher innate resistance to anti-cancer drugs compared to 2D cultures, more closely mimicking the in vivo scenario.^{www} This discrepancy contributes to a substantial translational gap, where drugs that show promise in 2D screens frequently fail in more complex preclinical models or clinical trials.^{www} Therefore, understanding how 2D culture conditions might specifically impact importin alpha expression and function is crucial for enhancing the predictive power of in vitro models and ultimately improving patient outcomes.

Part 2. Importin Alpha: Structure, Function, and Aberrant Expression in Cancer

2.1. Overview of Importin Alpha (Karyopherin Alpha) and its Role in Nuclear Import

Importin alpha, also known as Karyopherin alpha (KPNA), functions as a crucial adaptor protein within the classical nuclear import pathway, mediating the translocation of proteins from the cytoplasm into the nucleus.^{www} Its primary function involves the recognition and binding to specific nuclear localization signal (NLS) sequences, which are typically short sequences rich in basic amino acids, found on cargo proteins destined for nuclear entry.^{www} Following this recognition, importin alpha forms a ternary complex with importin beta (karyopherin beta 1) and the NLS-containing cargo protein.^{www} Importin beta then facilitates the passage of this entire complex through the nuclear pore complex (NPC), which serves as the gateway embedded in the nuclear envelope, into the nucleus.^{www}

Structurally, importin alpha is characterized by three functionally distinct domains: an N-terminal Importin-Beta Binding (IBB) domain, a central

Armadillo (ARM) domain, and an exportin CAS binding domain at the C-terminus.^{www} The IBB domain is responsible for the interaction with importin beta.^{www} The majority of the protein is composed of ten tandem ARM repeats, which form the NLS-binding cleft containing two binding sites, allowing a single importin alpha molecule to interact with either two monopartite NLS-containing proteins or a single bipartite NLS protein.^{www} An important auto-inhibitory mechanism exists within importin alpha: the IBB domain contains basic amino acid residues structurally similar to NLS sequences, enabling it to fold inward and occupy the NLS binding sites when importin beta is not associated. This mechanism prevents importin alpha from binding NLS-containing proteins prematurely, ensuring that cargo binding occurs only when all necessary import machinery is available, thereby increasing NLS affinity in the presence of importin beta.^{www}

Once the ternary complex reaches the nucleus, it dissociates due to the binding of importin beta to Ran-GTP, a small GTPase that is highly concentrated in the nucleus.^{www} Following cargo delivery, free importin alpha is recycled back to the cytoplasm via an export complex formed with exportin CAS and Ran-GTP, with the hydrolysis of GTP facilitating its dissociation and release into the cytoplasm.^{www} While nuclear import constitutes the primary functional role of importin alpha proteins, they have also been implicated in a diverse array of other crucial cellular functions, including gametogenesis, embryonic development, heat shock response, protein degradation (e.g., Taspase1), and even viral pathogen infection (e.g., influenza A virus, SIV).^{www}

2.2. Key Importin Alpha Isoforms and their Involvement in Carcinogenesis

The importin alpha gene family has undergone expansion in higher eukaryotes, with humans encoding seven distinct paralogs.^{www} These mammalian importin alpha paralogs often exhibit tissue-specific expression patterns and demonstrate preferential mediation of nuclear import for specific cargo proteins.^{www} A recurring theme in cancer biology is

the aberrant expression of various importin alpha isoforms, which significantly contributes to the malignant phenotype.

Karyopherin alpha 2 (KPNA2) has emerged as a particularly significant potential cancer biomarker due to its consistently elevated expression observed across a wide variety of cancer forms, including prostate cancer, liver cancer, bladder cancer, brain cancer, renal cell carcinoma (RCC), and neuroblastoma.^{www} This aberrant high level of KPNA2 is frequently associated with adverse patient characteristics, such as poor survival rates, increased tumor aggressiveness, and resistance to therapeutic interventions.^{www} Cellular studies have further substantiated KPNA2's direct role in malignant transformation, demonstrating that its deregulation affects cancer cell proliferation, migration, and invasion.^{www}

KPNA2's oncogenic influence is largely attributed to its crucial role in translocating various cancer-associated proteins into the nucleus. These include cell-cycle regulators like Chk2, DNA repair proteins such as BRCA1 and NBS1, and key transcription factors like the androgen receptor (AR), p53, and c-Myc.^{www} Overexpression of KPNA2 has been shown to correlate with increased nuclear import of Chk2 and direct interaction with BRCA1.^{www} This suggests that KPNA2 functions as a cargo-specific oncogenic amplifier. It is not merely enabling transport; its overexpression actively increases the nuclear concentration of factors that drive cell cycle progression, facilitate DNA repair (which can contribute to drug resistance), and promote hormone-dependent growth. This positions importin alpha as a critical node capable of modulating multiple cancer hallmarks simultaneously.

The regulation of KPNA2 expression itself can be influenced by E2F transcription factors, suggesting a potential positive feedback loop where elevated activity of E2Fs may lead to increased expression of KPNA2, which in turn boosts the nuclear amounts of E2Fs.^{www} Such positive feedback loops represent a critical vulnerability for therapeutic intervention. Disrupting KPNA2 could not only

reduce the nuclear import of its direct oncogenic cargoes but also break this self-sustaining mechanism of dysregulation, potentially leading to a more profound and sustained anti-cancer effect. KPNA2 expression also correlates with human papillomavirus (HPV) infection in tongue squamous cell carcinoma (TSCC) and is involved in cell differentiation and cancer-related pathways such as Cell Cycle, Mitotic G1 phase, G1/S transition, DNA Repair, and TP53 signaling.^{www}

Another isoform, Karyopherin- α 3 (KPNA3), has also been identified as significantly upregulated in certain cancer cells, particularly triple-negative breast cancer (TNBC).^{www} Knockdown of KPNA3 has been shown to suppress cell proliferation and metastasis in TNBC cell lines.^{www} KPNA3 promotes epithelial-mesenchymal transition (EMT) through the regulation of numerous EMT-related genes, including downregulation of GATA3 and E-cadherin and upregulation of HAS2.^{www} Furthermore, KPNA3-mediated EMT and metastasis can occur via independent signaling pathways involving TGF- β and AKT.^{www}

Karyopherin alpha 7 (KPNA7), the newest member of the importin alpha family, is highly expressed during embryogenesis but notably re-expressed in cancer cells, with its depletion inducing mitotic defects and nuclear deformation.^{www} Importin α 5 (KPNA1) has also been studied, with reduced expression in hippocampal neurons decreasing anxiety in mice by influencing MeCP2 nuclear localization and sphingosine kinase 1 (Sphk1) expression.^{www} While most importin alphas function as heterodimers with importin beta, with auto-inhibition by the IBB domain^{www}, it is important to note that importin- α 8 notably lacks an IBB domain.^{www} This structural difference suggests it may lack both importin-beta binding capacity and the conserved auto-inhibitory mechanisms, highlighting that generalizations about "importin alpha" function must be made with caution, as specific isoforms may possess unique mechanisms or roles, particularly in disease contexts. This nuance is crucial for targeted research and therapeutic development.

Table 1. Overview of Key Importin Alpha Isoforms and Their Documented Roles in Cancer

Importin Alpha Isoform (KPNA #)	Associated Cancer Type(s)	Reported Role/Function in Cancer	Key Cargoes/Pathways (if specified)
Importin α1 (KPNA2)	Prostate, Liver, Bladder, Brain, Renal Cell Carcinoma (RCC), Neuroblastoma, Tongue Squamous Cell Carcinoma (TSCC)	Potential biomarker for poor prognosis, associated with increased tumor aggressiveness and therapeutic resistance; promotes proliferation, migration, invasion, malignant transformation	Chk2, BRCA1, NBS1, Androgen Receptor (AR), p53, c-Myc, E2F, MRN complex, NPM, Cell Cycle, DNA Repair, TP53 signaling
Importin α4 (KPNA3)	Triple-Negative Breast Cancer (TNBC)	Significantly upregulated; knockdown suppresses cell proliferation and metastasis; promotes Epithelial-Mesenchymal Transition (EMT)	GATA3, E-cadherin, HAS2, TGF- β , AKT signaling pathways
Importin α5 (KPNA1)	Hippocampal neurons (non-cancer context, but highlights functional role)	Regulates anxiety; influences MeCP2 nuclear localization and Sphingosine Kinase 1 (Sphk1) expression	MeCP2, Sphk1
Importin α8 (KPNA7)	Cancer cells (re-expressed from embryonic levels)	Depletion induces mitotic defects and nuclear deformation	Not specified

Part 3. Limitations of 2D Cell Culture in Mimicking the Tumor Microenvironment

3.1. Fundamental Differences Between 2D and In Vivo/3D Conditions

The vast majority of cancer biology research relies heavily on two-dimensional (2D) cell cultures, where cells are grown as a monolayer, typically attached to a flat, rigid substrate made of plastic or glass.^{www} While these models offer undeniable advantages in terms of simplicity, low cost, and ease of performing functional tests ^{www}, they possess numerous fundamental limitations that severely compromise their ability to accurately mimic the complex and dynamic in vivo tumor microenviron-

ment (TME).^{www}

A primary drawback is the inherent failure of 2D cultured cells to recapitulate the natural three-dimensional (3D) architecture characteristic of tissues or tumors.^{www} In the physiological setting, cells are embedded within a complex 3D extracellular matrix (ECM) that provides crucial mechanical support and intricate biochemical cues.^{www} In stark contrast, 2D cultures largely lack or severely disturb essential cell-cell and cell-extracellular environment interactions.^{www} These interactions are not merely structural; they are vital for regulating critical cellular processes such as differentiation, proliferation, vitality, precise gene and protein expression, and appropriate responsiveness to stimuli.^{www} Upon isolation from native tissue and subsequent transfer to 2D culture conditions, cancer cells undergo significant alterations in their morphology and mode of cell division.^{www} They

frequently lose their diverse phenotype and cellular polarity, which can profoundly impact their intrinsic function, internal cellular organization, secretory processes, and cell signaling networks.^{www} Furthermore, 2D monolayer cultures provide cells with unlimited and uniform access to essential medium ingredients, including oxygen, nutrients, metabolites, and signaling molecules.^{www} This stands in sharp contrast to the heterogeneous in vivo tumor mass, where nutrient and oxygen availability is highly variable due to the natural architecture and often poor vascularization, leading to the formation of gradients and hypoxic regions.^{www} The absence of a complex tumor microenvironment, including various interacting cell types and specialized niches required by cancer-initiating cells, represents another significant limitation of simplified monoculture 2D systems.^{www}

This environment, so drastically different from the in vivo context, can exert an unnatural selective pressure. Cells that thrive and are preferentially selected for growth in 2D culture might possess different adaptive mechanisms or express different genes compared to those that flourish in a 3D, more physiological context. This selection process can lead to a skewed representation of cancer cell biology relevant to the human body, potentially impacting the expression of genes involved in cellular adaptation, including critical nuclear transport proteins like importin alpha.

In response to these pervasive limitations, three-dimensional (3D) culture methods have gained considerable popularity in recent years, as they are demonstrably better able to mimic the natural tumor mass and in vivo conditions.^{www} 3D models facilitate proper cell-cell and cell-extracellular environment interactions, preserve original cell morphology, polarization, genetic profile, and cellular heterogeneity, and crucially, create more realistic nutrient and oxygen gradients, thereby offering a more physiologically relevant experimental platform.^{www}

3.2. Impact of 2D Culture on Global Gene Expression and Cellular Phenotypes in Cancer Cells

The altered environment inherent in 2D cell cultures has a profound and pervasive impact on the global gene expression profiles, alternative splicing patterns, cellular topology, and biochemistry of cells.^{www} This leads to significant differences in cellular behavior and responses when compared to 3D cultures or authentic in vivo conditions. For instance, cells cultured in 3D exhibit distinct gene expression levels compared to their 2D counterparts, with 3D cultures more accurately reflecting the complex 3D tissue architecture and shaping cellular responses according to physiological microenvironmental cues.^{www}

These fundamental changes manifest in various hallmarks of cancer:

- Drug Resistance:** A consistent observation is that 3D cancer cell cultures demonstrate a higher innate resistance to anti-cancer drugs, such as neratinib and docetaxel, when compared to 2D cultures.^{www} This increased resistance is partly attributed to altered expression of receptor proteins, upregulation of drug transporters (e.g., P-glycoprotein), and increased activity of drug-metabolizing enzymes (e.g., CYP3A4) in 3D models.^{www} The exaggerated drug effects frequently observed in 2D models often provide misleading data regarding in vivo responses, contributing to the high failure rate of experimental drugs in clinical trials.^{www}
- Cell Viability and Proliferation:** Cell viability can be substantially lower in 3D cultures compared to 2D cultures.^{www} While 2D cultures tend to favor a predominantly proliferative cell population with unlimited access to nutrients, 3D cultures more accurately represent in vivo tumor heterogeneity by containing cells at various stages, including proliferation, quiescence, apoptosis, hypoxia, and necrosis.^{www}
- Morphology and Phenotype:** Cells grown in 2D cultures often lose some of their natural functional abilities, which can frequently be restored simply by growing them in 3D again.^{www} This includes significant changes in cell shape, loss of polarity, and the overall loss of a diverse

phenotype.^{www} For example, normal mammary epithelial cells, when cultured in 2D, lose their polarity and exhibit plasticity similar to malignant cells found in vivo. In contrast, when cultured in 3D, these cells self-organize into polarized acinar structures that closely resemble their physiological counterparts.^{www} Malignant cells in 3D form proliferating, disorganized aggregates without a hollow lumen, thereby mimicking in vivo tumor characteristics more closely than 2D monolayers.^{www}

The consistent observation that 2D systems change gene expression, splicing, topology, and

biochemistry ^{www} suggests a deeper, potentially epigenetic, reprogramming of the cell's state. If the fundamental regulatory mechanisms of gene expression are altered, then the entire cellular machinery, including the expression and function of nuclear transport proteins, would inevitably be affected. This implies that 2D culture does not merely "fail to mimic" the in vivo environment but actively *reprograms* cells in a manner that can make them fundamentally different entities from their in vivo counterparts, leading to a cascade of downstream effects on protein expression and function, including those of importin alpha.

Table 2. Comparative Analysis of 2D vs. 3D Cell Culture Models in Cancer Research

Feature	2D Culture Characteristics	3D Culture Characteristics
Tissue Architecture	Flat monolayer; does not mimic natural tissue structure ^{www}	Mimics in vivo 3D structure and organization ^{www}
Cell-Cell Interactions	Deprived or disturbed ^{www}	Proper and enhanced interactions; environmental "niches" created ^{www}
Cell-ECM Interactions	Deprived or disturbed ^{www}	Proper and enhanced interactions ^{www}
Cell Morphology	Altered, flattened; loss of diverse phenotype ^{www}	Preserved, in vivo-like shape and original phenotype ^{www}
Cell Polarity	Lost ^{www}	Preserved ^{www}
Nutrient/Oxygen Gradients	Uniform and unlimited access ^{www}	Variable availability, mimicking in vivo gradients (e.g., hypoxia) ^{www}
Gene Expression/Splicing	Altered and simplified; changes in gene expression and splicing ^{www}	More in vivo-like and complex; distinct gene expression levels ^{www}
Drug Response	Exaggerated sensitivity; unpredictable data compared to in vivo ^{www}	Increased resistance; better mimicry of in vivo drug response ^{www}
Heterogeneity	Monoculture; reduced heterogeneity; lack of TME niches ^{www}	Preserves tumor heterogeneity; creates environmental "niches" ^{www}
Cost/Simplicity	Low cost, simple maintenance, easy to interpret ^{www}	Higher cost, more complex to carry out and interpret ^{www}

Part 4. Mechanotransduction: A Bridge Between Physical Environment and Nuclear Regulation

The physical properties of the tumor microenvironment (TME), particularly the mechanical stiffness of the extracellular matrix (ECM), are increasingly recognized as critical regulators of cancer cell

behavior and gene expression.^{www}

Mechanotransduction is the intricate process by which cells sense and convert these mechanical inputs from their environment into biochemical signals, influencing a wide array of cellular functions, including proliferation, migration, differentiation, and apoptosis.^{www}

4.1. Extracellular Matrix (ECM) Stiffness and its Influence on Cancer Cell Behavior

Cancerous tissues typically exhibit significantly greater stiffness than their normal or adjacent counterparts, a hallmark feature of tumor progression.^{www} This increased stiffness results from excessive ECM synthesis, deposition, and crosslinking, often orchestrated by cancer-associated fibroblasts (CAFs), and provides potent mechanical cues that profoundly influence malignant phenotypes.^{www}

The impact of matrix stiffness extends across various cancer hallmarks:

- **Promotion of Malignancy:** Stiffer matrices promote numerous aspects of cancer progression, including enhanced proliferation, increased migration and invasion, angiogenesis, epithelial-mesenchymal transition (EMT), immune evasion, maintenance of stemness, metabolic reprogramming, and the development of therapeutic resistance.^{www} For instance, increased stiffness has been shown to reduce the sensitivity of tumor cells to anti-tumor drugs.^{www}
- **Alterations in Gene Expression:** Substrate stiffness directly modulates gene expression in cancer cells.^{www} Stiff substrates have been shown to increase the expression of specific receptors like CXCR4 and EGFR in triple-negative breast cancer cells^{www} and promote the expression of angiogenesis-related factors such as VEGFA, HIF-1 α , and TGF- β 1.^{www} They can also drive EMT by upregulating N-cadherin and vimentin and activating the TGF- β 1-induced Smad pathway.^{www} Hypoxia, a common feature of the TME often induced by

CAFs, is also a key inducer of ECM stiffness and degradation, creating a complex interplay of biophysical and biochemical cues.^{www}

The default 2D culture environment, typically comprising rigid polystyrene plastic, imposes an extreme mechanical stimulus a stiffness that is orders of magnitude higher than most physiological tissues and is only found in highly fibrotic tumors.^{www}

This constant, non-physiological mechanical input would continuously activate mechanotransduction pathways, leading to a sustained and potentially aberrant signaling state within cancer cells. Given that importin alpha expression is known to be dysregulated in cancer, and mechanotransduction profoundly influences gene expression, it is highly probable that this artificial stiffness in 2D culture directly contributes to the dysregulation of importin alpha expression, either by upregulating or downregulating specific isoforms as cells attempt to adapt to this unnatural environment. This represents a direct causal link inferred from the general principles of cellular mechanobiology.

4.2. Nuclear Mechanics, Cytoskeletal Tension, and Their Role in Regulating Nuclear Transport

The cell nucleus is not merely a passive container for genetic material but also acts as a crucial mechanosensor that experiences and actively responds to applied forces.^{www} Mechanical forces transmitted through the cell, particularly those arising from cytoskeletal tension, can directly alter nuclear shape, position within the cell, and overall nuclear function, including gene expression.^{www} The mechanical properties of the nucleus and its intricate connection to the cytoskeleton play a major role in cancer metastasis, as deformation of the large and relatively stiff nucleus can facilitate cancer cell invasion through dense tissues.^{www}

- **Force Transmission to the Nucleus:** Integrins, which link the cell to the ECM, and focal adhesions, large protein complexes connecting the cell cytoskeleton to integrins, are crucial for sensing and transmitting mechanical cues from the ECM.^{www} These forces are then relayed

through the cytoskeleton, ultimately reaching the nucleus via structures like the Linker of Nucleoskeleton and Cytoskeleton (LINC) complex.^{www} Cytoskeletal components, including actin, myosin, and intermediate filaments, undergo dynamic remodeling that is critical for cell migration and invasion in cancer.^{www}

- **Nuclear Deformation and Transport:** Cells cultured on stiff 2D substrates exert high intracellular force, which leads to mechanical deformation of their nuclei.^{www} This nuclear deformation (ND) plays a crucial role in the transport of mechanoresponsive proteins, such as Yes Associated Protein (YAP), from the cytoplasm to the nucleus.^{www} Studies indicate that perinuclear force, rather than the total cellular force, strongly correlates with both ND and nuclear YAP localization, suggesting that only a specific fraction of cellular force effectively contributes to nuclear deformation and subsequent nuclear transport.^{www} Alterations in nuclear envelope morphology, such as the frequent invaginations observed in cancer cells, can also affect the overall efficiency of nucleocytoplasmic transport.^{www} Furthermore, the depletion of certain importin alpha isoforms, such as KPNA7, has been shown to induce mitotic defects and nuclear deformation in cancer cells, suggesting a direct link between importins and nuclear mechanics.^{www}
- **Stress Responses and Nuclear Importin Alpha:** Various cellular stresses, including oxidative stress and heat shock, can induce a collapse of the Ran gradient, a key regulator of nuclear transport, leading to the accumulation of importin alpha (specifically importin $\alpha 2$) in the nucleus and a subsequent block of nuclear protein import.^{www} This nuclear-localized importin alpha can then influence gene expression.^{www} Interestingly, importin alpha has also been observed to be highly concentrated in micronuclei in cultured human cancer cells, suggesting a non-canonical function in genome instability.^{www}

If 2D culture conditions impose unnatural mechani-

cal stresses leading to altered nuclear mechanics and deformation, this could fundamentally alter the nuclear import/export machinery and its efficiency. The observation that KPNA7 depletion induces nuclear deformation^{www} and that importin alpha accumulates in micronuclei^{www} points to a direct connection between importins and nuclear mechanics. This implies that targeting nuclear mechanics or the proteins that link the cytoskeleton to the nucleus (like the LINC complex^{www}) could represent a novel therapeutic strategy to indirectly dysregulate nuclear transport and, by extension, importin alpha activity, offering a new avenue beyond direct protein inhibition.

4.3. Key Mechanosensitive Signaling Pathways and Their Link to Nuclear Function

Mechanotransduction pathways serve as the crucial mediators that translate mechanical cues from the cellular environment into changes in gene expression, often relying on the nuclear import of mechanoresponsive transcriptional regulators.^{www}

- **Hippo/YAP/TAZ Pathway:** The Hippo pathway, with its core transcriptional regulators YAP (Yes-associated protein) and TAZ (transcriptional coactivator with PDZ-binding motif), is a prime example of a mechanosensitive pathway that is critically involved in cancer progression.^{www} YAP/TAZ are activated by a stiff extracellular matrix and promote malignant phenotypes in both cancer cells and associated stromal cells.^{www} Under normal physiological conditions, activation of the Hippo pathway leads to the phosphorylation and subsequent cytoplasmic sequestration of YAP and TAZ, thereby inhibiting their transcriptional activity.^{www} However, when the pathway is inhibited, often by mechanical cues such as increased matrix stiffness, YAP and TAZ translocate to the nucleus, where they interact with the TEAD family of transcription factors to activate genes that promote cell proliferation and inhibit apoptosis.^{www}
- **YAP and Importin 7 (Imp7):** YAP has been identified as a mechanoresponsive cargo of

Importin 7 (Imp7), a nuclear transport receptor.^{www} Intriguingly, YAP appears to govern the mechanoreponse of Imp7 by forming a YAP/Imp7 complex that responds to mechanical cues through the Hippo kinases (MST1/2).^{www} Furthermore, YAP behaves as a dominant cargo of Imp7, actively restricting the binding and nuclear translocation of other Imp7 cargoes, such as Smad3 and Erk2.^{www} This suggests that mechanical cues, such as those imposed by stiff 2D culture substrates, can indirectly regulate the nuclear import process not just by altering overall transport efficiency, but by actively re-prioritizing which proteins get imported. This "mechanically-driven nuclear transport re-prioritization" could lead to a dysregulated nuclear proteome, where oncogenic factors like YAP are preferentially imported, while other crucial factors are excluded, thereby creating signaling crosstalk and contributing to the malignant phenotype.

- **Other Mechanosensitive Pathways:** ECM stiffness also activates other significant signaling pathways, including Integrin/FAK, AKT, β -catenin, and PI3K, and can inhibit tumor suppressor genes like PTEN and GSK3 α/β , all of which contribute to tumorigenesis.^{www}

The consistent narrative emerging from these observations is one of profound interconnectedness: the physical environment, particularly ECM stiffness, dictates cellular mechanics, including cytoskeletal tension and nuclear deformation. These mechanical cues, in turn, activate specific mechanosensitive signaling pathways, most notably the Hippo/YAP/TAZ pathway, which ultimately influences gene expression and nuclear transport. This chain of causality provides a robust framework for understanding how the artificial mechanical properties of 2D culture could lead to dysregulation of nuclear transport components, including importin alpha.

Table 3. Impact of Biophysical Cues on Cancer Cell Behavior and Gene Expression in 2D Culture

<i>Biophysical Cue</i>	<i>Effect on Cancer Cell Behavior/Phenotype</i>	<i>Effect on Gene Expression/Signaling</i>
ECM Stiffness (High)	Increased proliferation, migration, invasion, drug resistance, EMT, altered morphology ^{www}	Upregulation of CXCR4, EGFR, VEGFA, HIF-1 α , TGF- β 1; activation of Akt, ERK, FAK, PI3K, AKT, β -catenin; increased YAP/TAZ nuclear translocation; altered splicing ^{www}
Nuclear Deformation	Facilitates cancer cell invasion; linked to mitotic defects ^{www}	Crucial for YAP nuclear transport; affects nucleocytoplasmic localization of transcriptional regulators; influences gene expression ^{www}
Cytoskeletal Tension	Critical for cell migration and invasion; influences cell shape and polarity ^{www}	Regulates YAP/TAZ phosphorylation and nuclear translocation; affects nuclear receptors; influences gene expression ^{www}
Lack of 3D Cell-Cell/ECM Interactions	Disturbed differentiation, proliferation, vitality; altered morphology and polarity; loss of diverse phenotype ^{www}	Impaired gene expression and protein responsiveness to stimuli; leads to compensatory/maladaptive gene expression programs ^{www}

2D Cancer Cell Cultures

Part 5. Investigating the Dysregulation of Importin Alpha Expression in

The central question of how 2D culture conditions specifically dysregulate importin alpha expression

in cancer cell lines necessitates a careful examination of both direct empirical evidence and a robust exploration of indirect mechanistic pathways.

5.1. Direct Evidence: Scrutiny of Studies Directly Linking 2D Culture Conditions to Importin Alpha Expression Changes in Cancer Cells

Upon comprehensive review of the available research, direct evidence explicitly detailing how variations within 2D culture conditions (e.g., differences in substrate stiffness, cell densities, or media compositions *within a 2D context*) lead to the dysregulation of importin alpha *expression levels in cancer cell lines* is notably limited.

One observation indicates that importin $\alpha 1$ (KPNA2) localizes to the cell surface in several cancer cell lines cultured in vitro and is detectable in their culture medium, where it enhances FGF1 signaling and accelerates proliferation.^{www} This is an important finding regarding importin alpha's presence and function in cultured cancer cells, including its non-canonical roles. However, this study does not compare different 2D culture conditions or explicitly link the 2D environment to *dysregulation of its expression levels* when compared to a more physiological state.

Several studies extensively compare 2D and 3D cell cultures, noting profound differences in cell morphology, viability, drug resistance, and global gene expression.^{www} While these investigations highlight that "the 2D system changes the gene expression and splicing, topology and biochemistry of the cell"^{www}, they do not specifically mention importin alpha or other nuclear transport proteins as being among the dysregulated genes or proteins in their comparative analyses.^{www}

Furthermore, studies on non-cancer cells or general stress responses, while informative, do not directly address the query. For instance, one article discusses how substrate stiffness in 2D culture affects importin alpha expression in *mouse embryonic fibroblasts (MEFs)*, noting a ~50% increase in importin $\alpha 3$ and a ~40% decrease in importin $\alpha 1$ with stiffness.^{www} However, it explicitly states this

discussion does not extend to *cancer cells*.^{www} Another study demonstrates that various cellular stresses, such as oxidative stress and heat shock, induce nuclear accumulation of importin alpha (specifically importin $\alpha 2$) in HeLa cells (a cancer cell line) by collapsing the Ran gradient, which in turn influences gene expression.^{www} While this illustrates importin alpha's sensitivity to stress in cancer cells, it does not attribute the dysregulation directly to the general 2D culture conditions or the mechanical stress inherent to 2D.^{www} Similarly, a study on cancer-associated fibroblasts (CAFs) discusses YAP dynamics in 2D versus 3D but explicitly states it does not discuss importin alpha.^{www}

Therefore, while importin alpha is well-established as aberrantly expressed in cancer^{www} and 2D culture profoundly alters global gene expression^{www}, direct empirical evidence specifically linking the *conditions of 2D culture* to the *dysregulation of importin alpha expression in cancer cell lines* is not explicitly provided in the available literature. This suggests a significant knowledge gap that necessitates inference from indirect mechanisms. It is highly improbable that the nuclear transport machinery, which is so central to cell function and known to be dysregulated in cancer, would be immune to the profound environmental changes imposed by 2D culture. This represents an implicit dysregulation that warrants further investigation. The absence of direct evidence, despite extensive literature on both 2D culture limitations and importin alpha in cancer, also points to a methodological blind spot. Researchers often focus on the *consequences* of 2D culture (e.g., drug resistance, altered proliferation) or the *role* of importin alpha in cancer, without explicitly investigating the *interplay* between the 2D environment and importin alpha expression. This highlights a critical area for future research: direct comparative studies of importin alpha expression, localization, and activity in cancer cells cultured under varying 2D conditions and contrasted with 3D models.

5.2. Indirect Mechanisms: How Altered Mechanical Cues in 2D Culture May Influence Importin Alpha Expression and Function

Despite the limited direct evidence, a strong argument can be made for indirect mechanisms through which 2D culture conditions likely influence importin alpha expression and function, primarily via the altered mechanical cues inherent to the 2D environment.

- **Altered Mechanical Environment and ECM Stiffness:** 2D cell culture substrates, typically made of polystyrene plastic, are unnaturally stiff, often orders of magnitude stiffer than most physiological tissues and tumors.²⁴ This high stiffness is a major biophysical property that continuously triggers mechanotransduction pathways.²⁴ This constant, non-physiological mechanical input would lead to a sustained and potentially aberrant signaling state. Given that importin alpha expression is known to be dysregulated in cancer, and mechanotransduction influences gene expression, it is highly probable that this artificial stiffness in 2D culture directly contributes to the dysregulation of importin alpha expression, either by upregulating or downregulating specific isoforms as cells adapt to this unnatural environment.
- **Impact on Nuclear Mechanics and Transport:** Cells cultured on stiff 2D substrates exert high intracellular force, which leads to mechanical deformation of their nuclei.⁵² Nuclear mechanics and deformability are crucial for cell behavior, particularly in the context of cancer cell invasion.⁵⁰ Mechanical forces transmitted to cells, and specifically to their nuclei, affect the nucleocytoplasmic localization of transcriptional regulators.⁴⁹ Nuclear deformation plays a crucial role in the transport of proteins like YAP from the cytoplasm to the nucleus.⁵² Importin alpha's primary role is nuclear import, which occurs through nuclear pore complexes embedded in the nuclear envelope.¹ The nuclear envelope and its associated proteins are critical for nuclear mechanics and deformability.⁵⁰ If 2D culture conditions induce abnormal nuclear deformation ⁵², it is highly plausible that this physical stress directly impacts the integrity and function of nuclear pore complexes

and the efficiency of importin alpha-mediated transport. This could lead to altered subcellular localization of importin alpha itself (e.g., increased nuclear retention due to Ran gradient collapse under stress ⁵⁷) or changes in its ability to bind and transport cargo, thus functionally dysregulating it even if expression levels remain constant.

- **Mechanosensitive Signaling Pathways and Gene Expression:** The Hippo/YAP/TAZ pathway is highly sensitive to matrix stiffness, and its activation promotes malignant phenotypes and influences gene expression.²⁴ YAP/TAZ nuclear translocation is a key step in mechanotransduction.⁴⁷ YAP functions as a dominant cargo for Imp7, and its nuclear translocation in response to mechanical cues can restrict the import of other cargoes.⁵⁹ Given that YAP/TAZ are major transcriptional regulators influenced by stiffness ³⁰, and 2D cultures provide an artificially stiff environment, YAP/TAZ would likely be constitutively active and nuclear in many 2D cancer cell lines. This sustained activation could directly or indirectly regulate the gene expression of importin alpha isoforms. For instance, if YAP/TAZ promote the expression of genes involved in proliferation and malignancy, and importin alpha isoforms like KPNA2 and KPNA3 are also linked to these processes ⁴, a regulatory link is highly probable. The competition for importins ⁵⁹ also suggests that even if importin alpha expression is not directly altered, its *functional availability* for certain cargoes could be dysregulated due to the altered mechanosensitive signaling.
- **Loss of Cell-Cell and ECM Interactions:** 2D cultures deprive cells of proper cell-cell and cell-extracellular environment interactions, which are crucial for cell differentiation, proliferation, vitality, and gene expression.⁸ The absence of these physiological interactions in 2D removes critical regulatory cues that normally maintain cellular homeostasis and phenotype. This "loss of context" can lead to a compensatory or maladaptive gene expression program. Since importin alpha expression is tightly regulated and often aberrantly expressed in cancer, the lack

of appropriate cell-cell and ECM signaling in 2D could directly contribute to its dysregulation as cells attempt to adapt to this impoverished environment. This represents a broad, systemic effect that would likely cascade to key regulatory proteins like importin alpha.

- **Altered Nutrient/Oxygen Gradients (Hypoxia):** 2D monolayers have unlimited access to medium ingredients, unlike the variable nutrient and oxygen availability in vivo due to tumor architecture and hypoxia.⁸ Hypoxia is a significant factor in the TME, influencing gene expression and cancer progression.³² The absence of physiological oxygen and nutrient gradients in 2D cultures means that cells are not exposed to the hypoxic stress common in tumors. Hypoxia-inducible factors (HIFs) are stabilized at low oxygen levels and activate numerous downstream pathways, influencing proliferation, survival, and metastasis.³² While the available information does not directly link hypoxia to importin alpha expression, it is a known regulator of gene expression. Therefore, the lack of physiological hypoxia in 2D could prevent the activation of certain pathways that might normally regulate importin alpha expression in vivo, leading to a different expression profile compared to the TME. Conversely, the uniform nutrient access might also lead to distinct metabolic states that indirectly impact gene expression.

5.3. Implications for Importin Alpha Subcellular Localization and Activity in 2D Models

The indirect mechanisms discussed above have significant implications for not only importin alpha expression levels but also its subcellular localization and functional activity in 2D cancer cell models.

- **Altered Localization:** Changes in nuclear mechanics, integrity of the Ran gradient (especially under stress conditions), or altered binding partners due to mechanotransduction could affect the nuclear-cytoplasmic shuttling of importin alpha.^{www} For example, stress-induced

Ran gradient collapse leads to nuclear accumulation of importin alpha.^{www} If 2D culture itself induces cellular stress, perhaps due to the mechanical mismatch with the substrate, this could lead to non-physiological importin alpha localization, thereby functionally compromising its role. This represents a functional dysregulation beyond mere expression changes.

- **Modified Cargo Binding and Transport Efficiency:** The auto-inhibitory mechanism of importin alpha's IBB domain and its affinity for NLS sequences are tightly regulated by importin beta association.^{www} If 2D conditions alter the expression, availability, or post-translational modification of importin beta or other regulatory proteins, the cargo binding affinity and overall transport efficiency of importin alpha could be compromised. Furthermore, the mechanically-driven nuclear transport re-prioritization, where dominant cargoes like YAP can restrict the import of others ^{www}, suggests that even if importin alpha expression is not directly altered, its *effective* function in transporting specific oncogenic or tumor-suppressive cargoes could be severely dysregulated due to competition or altered signaling.
- **Surface Localization and Non-Canonical Functions:** The observation that importin α 1 (KPNA2) can localize to the cell surface in cancer cell lines in culture and affect FGF1 signaling ^{www} suggests an additional layer of complexity. This non-canonical localization for a protein primarily known for nuclear import might be influenced or even exaggerated by 2D culture conditions. Given that 2D culture significantly alters cell morphology, polarity, and interactions ^{www}, it is plausible that the artificial 2D environment could induce or exaggerate such non-canonical localizations and functions of importin alpha. This implies that 2D models might not just fail to mimic in vivo dysregulation, but could actively *create* novel, artificial dysregulations or functions that are not relevant to the in vivo tumor, potentially leading to misinterpretations in research.

Part 6. Conclusion and Future Research Directions

The comprehensive analysis indicates that while importin alpha proteins, particularly isoforms like KPNA2 and KPNA3, are frequently dysregulated and play critical roles in cancer progression in vivo, direct, explicit evidence demonstrating how standard 2D cell culture conditions *specifically* dysregulate their *expression levels* in cancer cell lines is not prominently featured in the provided literature. However, a robust framework of indirect mechanisms strongly suggests that such dysregulation is highly probable and multifaceted.

The inherent limitations of 2D cultures – including their unnatural stiffness, lack of physiological 3D architecture, disturbed cell-cell and ECM interactions, and uniform nutrient/oxygen access – profoundly alter global gene expression, cellular morphology, and mechanotransduction pathways in cancer cells. These alterations, particularly the activation of mechanosensitive pathways like Hippo/YAP/TAZ and the induction of nuclear deformation, create a biophysical and biochemical environment distinct from the in vivo TME. Given that importin alpha expression is known to be sensitive to cellular stress and is a key component of a mechanically-influenced nuclear transport system, it is highly plausible that 2D culture conditions indirectly lead to dysregulation of importin alpha expression, localization, and functional activity. This dysregulation may involve altered transcriptional regulation of importin alpha genes, changes in the efficiency of nuclear-cytoplasmic shuttling, or competitive binding for importin alpha by mechanosensitive cargoes. The absence of direct studies on importin alpha dysregulation specifically due to 2D culture conditions in cancer cells represents an implicit dysregulation and a methodological blind spot in current research.

Summary of Current Understanding and Identified Gaps:

- Importin alpha isoforms are crucial for nuclear transport and are often aberrantly expressed in various cancers, contributing significantly to

malignancy.

- 2D cell cultures drastically differ from the in vivo TME, leading to profound changes in global gene expression, cell behavior, and drug response, often yielding misleading results.
- Mechanotransduction, driven by ECM stiffness and nuclear mechanics, profoundly influences gene expression and nuclear transport, with pathways like Hippo/YAP/TAZ playing a central and re-prioritizing role.
- A key knowledge gap exists in direct, explicit studies comparing importin alpha expression and function in cancer cell lines across different 2D conditions (e.g., varying stiffness within 2D) or directly comparing 2D versus 3D cultures for importin alpha specifically. The existing evidence primarily infers dysregulation through broader changes in mechanotransduction and gene expression.

Recommendations for Future Research Directions:
To bridge this critical knowledge gap and enhance the physiological relevance of in vitro cancer research concerning importin alpha, future studies should focus on the following key areas:

- **Direct Comparative Studies:** Systematically compare the expression levels (mRNA and protein), subcellular localization, and functional activity of key importin alpha isoforms (e.g., KPNA2, KPNA3) in cancer cell lines cultured under various 2D conditions (e.g., different substrate stiffnesses, ECM coatings, cell densities). Crucially, these studies should be conducted in parallel with 3D models that better mimic the in vivo TME to provide a physiological benchmark.
- **Mechanistic Elucidation:** Investigate the specific mechanotransduction pathways (e.g., integrin-FAK, Hippo-YAP/TAZ, LINC complex signaling) that mediate the effects of 2D culture-induced mechanical cues on importin alpha gene expression and nuclear transport dynamics. This could involve targeted interventions using pharmacological inhibitors or genetic manipulation of these pathways in 2D cultures to understand their direct impact.
- **Cargo-Specific Analysis:** Beyond assessing

overall importin alpha expression, it is vital to evaluate how 2D culture conditions influence the nuclear import and localization of *specific* oncogenic or tumor-suppressive cargo proteins known to interact with importin alpha (e.g., Chk2, BRCA1, AR, p53, c-Myc, YAP). This would reveal the functional consequences of potential dysregulation on critical cancer-related pathways.

- **Multi-Omics Approaches:** Employ advanced multi-omics techniques, such as transcriptomics, proteomics, and spatial proteomics, to comprehensively map the changes in nuclear transport machinery components and their cargo profiles when cancer cells transition from 2D to 3D culture or are exposed to varying mechanical environments. This holistic approach can identify novel regulatory networks.

- **Integration with Clinical Data:** Correlate in vitro findings with data from more complex pre-clinical models, such as patient-derived organoids or xenograft models, and ultimately with clinical data on importin alpha expression and patient outcomes. This crucial step will ensure the translational relevance of in vitro discoveries.

By rigorously addressing these research avenues, a more precise and physiologically relevant understanding of how in vitro culture conditions influence importin alpha biology in cancer can be achieved. This enhanced understanding is essential for developing more accurate drug discovery platforms and ultimately, more effective therapeutic strategies for cancer patients.